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# ***A method for finding molecular signatures from gene expression data***

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# Introduction

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- *Molecular signatures* or *Gene expression signatures* are a key feature in many papers in cancer research. For instance, Alizadeh et al., 2000; Golub et al., 1999; Huang et al., 2002; Pomeroy et al. 2002; Ramaswamy et al., 2003; Rosenwald et al., 2002; Shipp et al., 2002; Yeoh et al., 2002.
- A possible definition: “(...) a group of genes expressed in a specific cell lineage or stage of differentiation or during particular biological response.” (Rosenwald et al., 2002, N. Eng. J. Med., 346, p. 1942)
- Often used as independent variables to model clinically relevant information (cancer vs. healthy, survival time, etc).
- Provide insight into biological mechanisms and processes and have potential diagnostic use.
- However, searching for molecular signatures often done using a very diverse and ad-hoc methodology.

## What we want:

- Find groups of genes [“group of genes” = “signature component”] so that genes within a group are tightly coexpressed, and the set of groups do a decent predictive job.
- Nice if a similar procedure can be applied to different types of dependent (phenotypic) data (e.g., class membership, survival data, expression of a relevant protein).
- Should help gain some understanding, not necessarily find **The** best predictor (flexibility to play around with trade-offs).

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- Signature components could have many genes.



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- **Tight co-expression:** We can use *Principal Components Analysis* to characterize a signature component. The first principal component of a PCA on the genes that belong to a signature component should capture a large fraction of the total variance of those genes.
- **Few signature components:** Add new components only if justified. (Is this always what we want to do?).
- **Signature components could have many genes:** Retain as many genes per component as possible.
- **Predictor:** Build a predictor using the signature components (1st PCs). **Diagonal Linear Discriminant Analysis** (*DLDA*).

- We have:  $\mathbf{Y}$  ( $n \times q$ ),  $\mathbf{X}$  ( $n \times p$ ),  $p \gg n$ .
- We want:  $\mathbf{Y}$ ,  $\mathbf{X}^*$  ( $n \times k$ ),  $k < n$ .
- $\mathbf{X} = [\mathbf{x}_1, \dots, \mathbf{x}_{pr1,1}, \mathbf{x}_{pr1,2}, \mathbf{x}_{pr1,3}, \dots, \mathbf{x}_{pr2,1}, \mathbf{x}_{pr2,2}, \dots]$
- $\mathbf{X}^* = [\mathbf{pr}_1, \mathbf{pr}_2, \dots, \mathbf{pr}_k]$ .
- $\mathbf{pr}_i$  is the  $i$ th signature component or profile, and it is the 1st PC of a PCA on genes  $\mathbf{x}_{pr_i,1}, \mathbf{x}_{pr_i,2}, \dots, \mathbf{x}_{pr_i,m_i}$ .
- Each gene belongs to either one (and only one) signature component or to none.
- $\mathbf{Y} = f(\mathbf{X}^*) + \epsilon$ .

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 $pred.error.model_i < last.pred.error - c_1 s.e. (c_1 = 1)$ .
- **Initial signature component**: all genes with abs. corr. with seed gene  $> r_{seed}$  (e.g.,  $r_{seed} = 0.65$ ).
  - These are the candidate genes to belong to that component.
  - But this initial signature component might not fulfill previous requirements (%var, predictive performance).
  - Examine if elimination of genes is needed.

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- Ensure that predictive accuracy cannot be improved by removing any gene from signature component:
  - For each gene,  $i$ , in current signature component:  $model_i = DLDA(previous.components + current.component_{-i})$ .
  - Eliminate gene  $i$  from signature component if (cross-validated)  $pred.error.model_i < last.pred.error - c_2 s.e.$  ( $c_2 = 1$ ).
  - Repeat until no gene is eliminated.

# Bootstrap

We use the bootstrap to assess stability of results and measure prediction error (.632+ rule).

- Take  $B$  ( $= 100$ ) bootstrap samples, and for each one run the above procedure.
- *Common genes*: genes that are returned in at least 20% of the samples.
- For each run, eliminate from the signature components those genes that are not in common genes to obtain “clean signature components”.
- *Consensus signature components* are obtained as the (most inclusive) union of all “clean signature components” with a non-zero intersection.

# Can we recover signatures?

- Simulation study.
- Generate signature data from a multivariate normal distribution.
- Correlation between genes within a signature component: 0.9. Between genes among signature components: 0. (i.e.,

$$\Sigma = \begin{bmatrix} a & \mathbf{0} & \mathbf{0} & \cdots & \mathbf{0} \\ \mathbf{0} & a & \mathbf{0} & \cdots & \mathbf{0} \\ \vdots & \vdots & \vdots & \vdots & \vdots \\ \mathbf{0} & \mathbf{0} & \mathbf{0} & \cdots & a \end{bmatrix},$$

$$a = \begin{bmatrix} 1 & 0.9 & \cdots & 0.9 \\ 0.9 & 1 & \cdots & 0.9 \\ \vdots & \vdots & \vdots & \vdots \end{bmatrix}$$

).

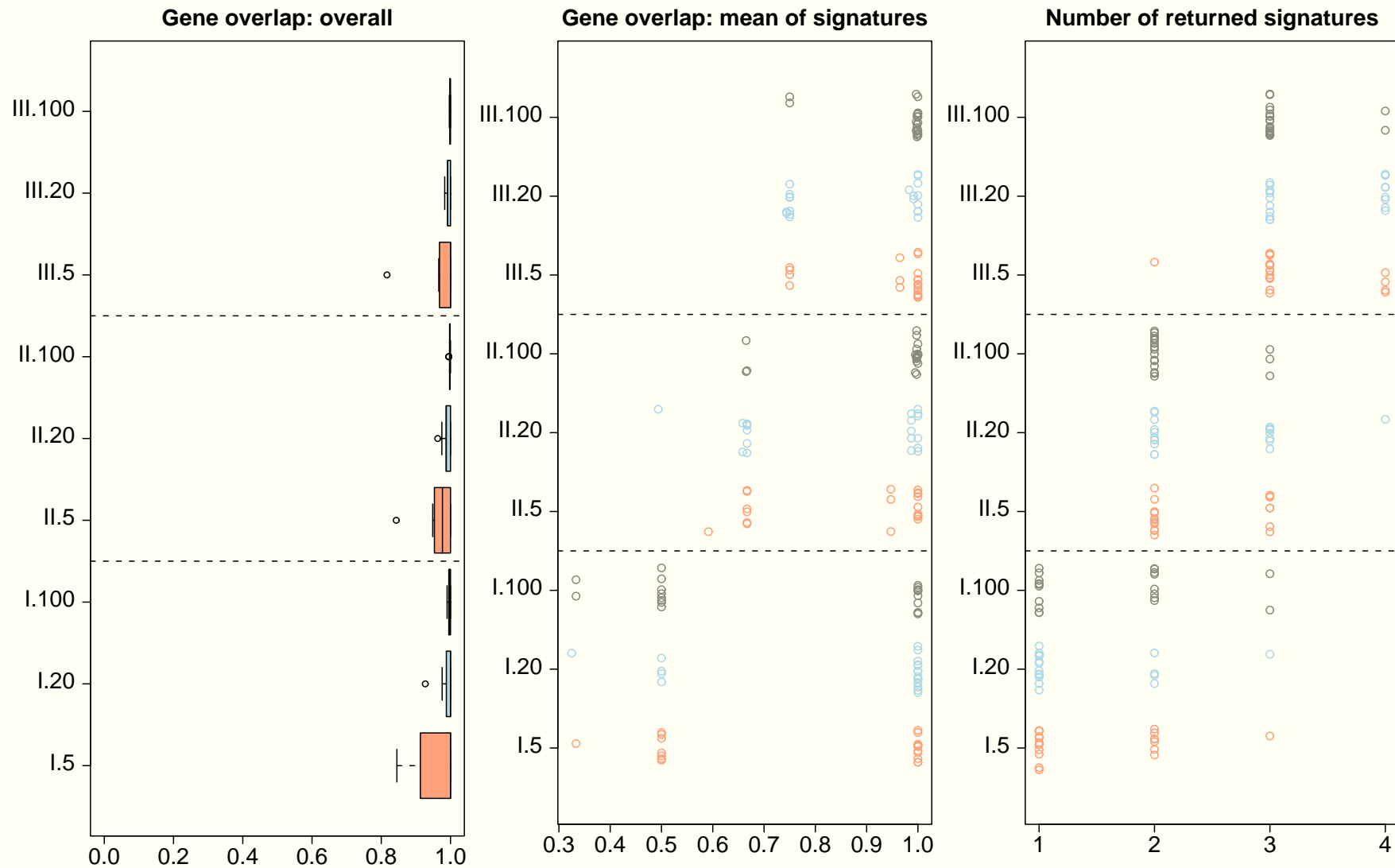


- Means of classes set so that:
  - unconditional prediction error rate of a DLDA with a gene from each signature component is approx. 5%;
  - each signature component has the same relevance in separation.
- Number of signature components: {1, 2, 3}.
- Number of classes: {2, 3, 4}.
- Number of genes per signature component: {5, 20, 100}.
- Add another 4000  $N(0, 1)$  variables to matrix of covariates.
- Number of subjects: 25 per class.
- Generate 20 data sets and run procedure.

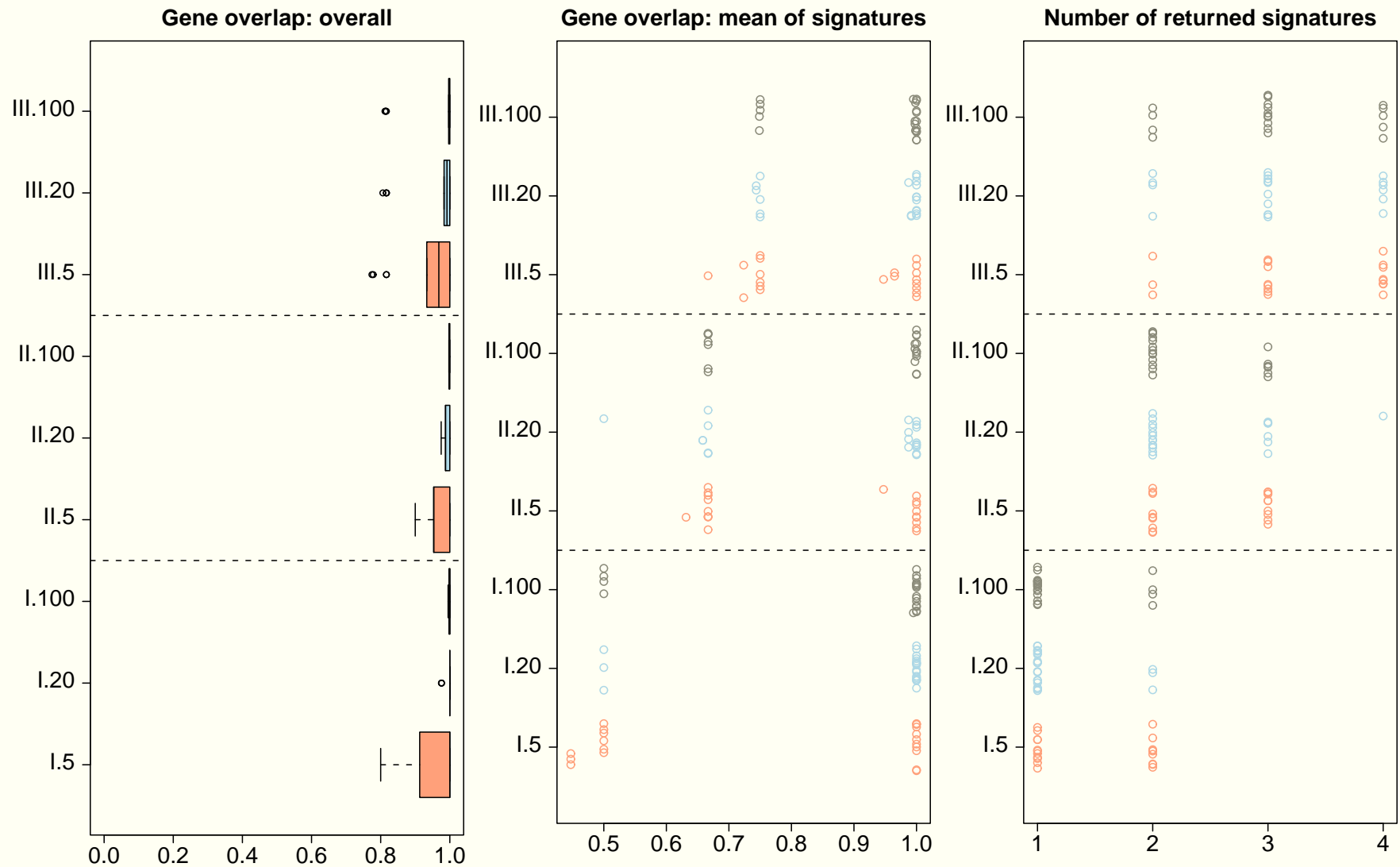
# Class means

- One signature component:
  - Two classes:  $\mu_1 = -1.65, \mu_2 = 1.65$ .
  - Three classes:  $\mu_1 = -3.58, \mu_2 = 0, \mu_3 = 3.58$ .
  - Four classes:  $\mu_1 = -3.7, \mu_2 = 0, \mu_3 = 3.7, \mu_4 = 7.4$ .
- Two signature components:
  - Two classes:  $\mu_1 = [-1.18, -1.18], \mu_2 = [1.18, 1.18]$ .
  - Three classes:  $\mu_1 = [0, 0], \mu_2 = [3.88\cos(15), 3.88\sin(15)], \mu_3 = [3.88\cos(75), 3.88\sin(75)]$ .
  - Four classes:  $\mu_1 = [1, 1], \mu_2 = [4.95, 1], \mu_3 = [1, 4.95], \mu_4 = [4.95, 4.95]$ .
- Three signature components:
  - Two classes:  $\mu_1 = [-0.98, -0.98, -0.98], \mu_2 = [0.98, 0.98, 0.98]$ .
  - Three classes:  $\mu_1 = [2.76, 0, 0], \mu_2 = [0, 2.76, 0], \mu_3 = [0, 0, 2.76]$ .
  - Four classes:  
 $\mu_1 = [2.96, 0, 0], \mu_2 = [0, 2.96, 0], \mu_3 = [0, 0, 2.96], \mu_4 = [2.96, 2.96, 2.96]$

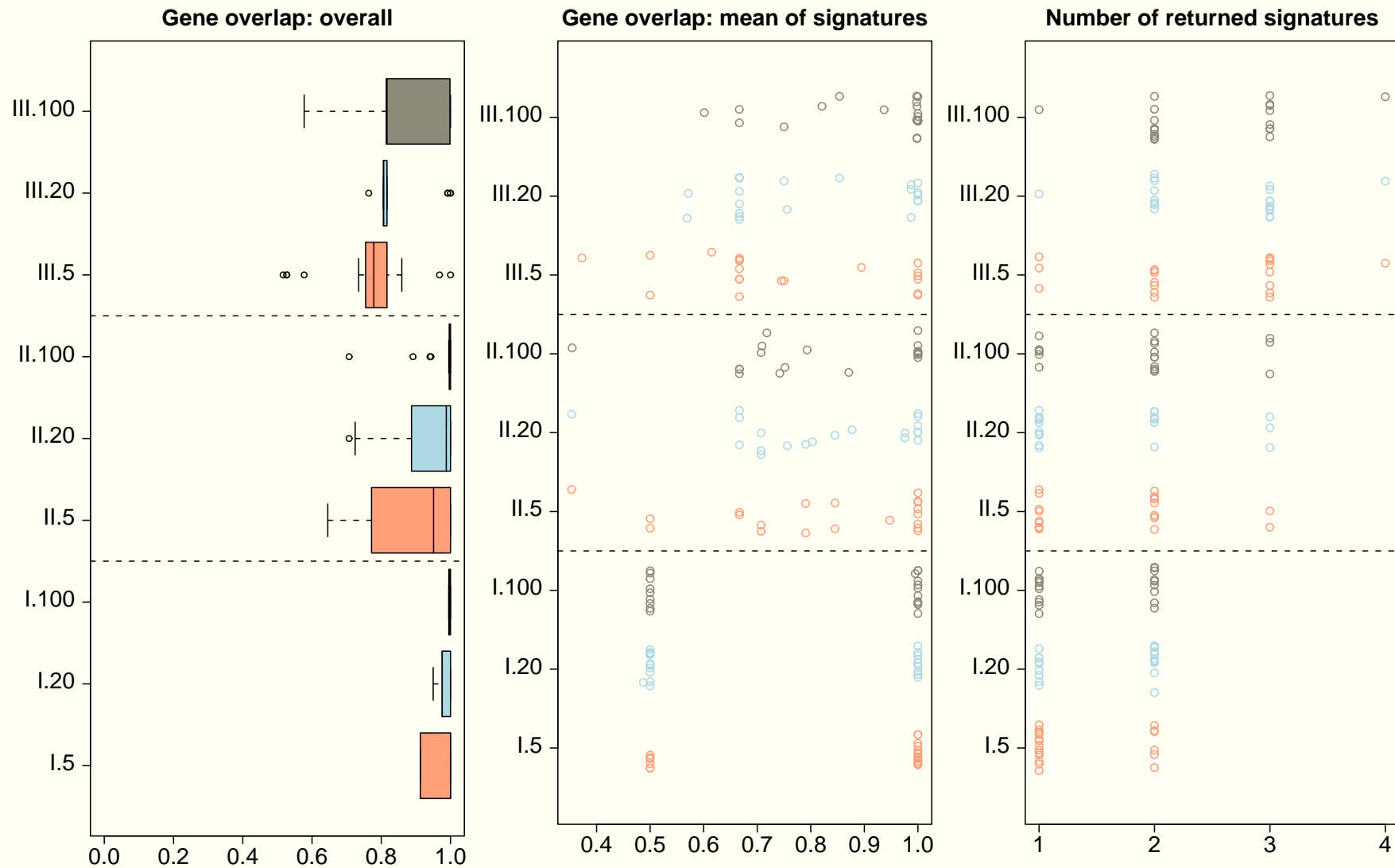
## Four classes



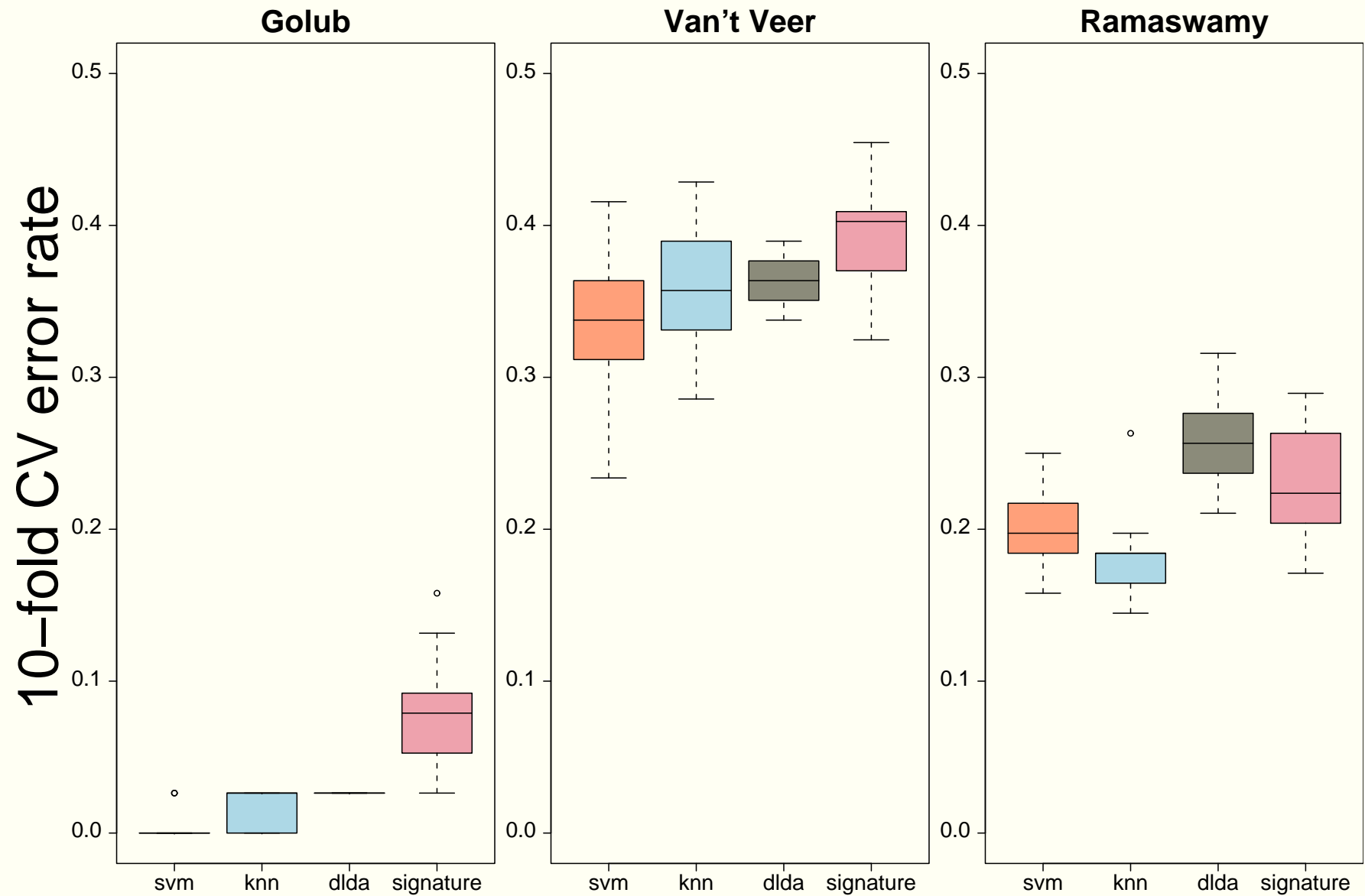
## Three classes



## Two classes



# Comparison with standard methods



# *Stability of results?*

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- Models on all data:
  - Golub: 1st PC > 85%: 1 comp. (1 gene); 1st PC > 75%: 1 comp. (6 genes); 1st PC > 70%: 2 comp. (11 and 19 genes).
  - van't Veer: 1 comp. (1 gene); 1 comp. (3 genes); 5 comp. (13 genes); ...
  - Ramaswamy: 1 comp. (1 gene); 2 comp. (10 and 1 genes); 2 comp. (2 genes); ...

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  - van't Veer: 1 comp. (1 gene); 1 comp. (3 genes); 5 comp. (13 genes); ...
  - Ramaswamy: 1 comp. (1 gene); 2 comp. (10 and 1 genes); 2 comp. (2 genes); ...
- Bootstrap:
  - 1st PC > 85% var:
    - Golub: 1 comp. with 19 genes;
    - van't Veer and Ramaswamy: no common genes;
  - 1st PC > 75% var:
    - Golub: 1 comp. with 48 genes;
    - van't Veer: no common genes;
    - Ramaswamy: 1 comp. of 2 genes;



# Discussion

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  - Appropriate threshold for % var., correlations, etc?
  - Entry of a component, given previous components?
  - Within-group heterogeneity?
  - PCA: between vs. within group patterns.

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Easily extended:

- Other classifiers (e.g., logistic regression, knn, svm).
- Other dependent variables: survival analysis.

## *Related to*

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- Partial Least Squares (and, to a lesser extent, Principal Components Regression).
- Factor analysis with oblique rotations to obtain clusters of variables (SAS's PROC VARCLUS).
- “Supergenes” or “metagenes” of West et al.
- ...

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# Conclusion

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- The logic of this method follows directly from what are considered biologically relevant signature characteristics.
- Results that are biologically relevant and interpretable and can complement other approaches.
- This method defines a framework that allows us to find signatures regardless of the type of dependent variable.
- Easy to implement and R code available.

# ***Acknowledgements***

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